

# Unlocking the Antioxidant Potential of Malaysian Traditional Salad (Ulam) as a *Halal* Natural Carotenoid Source

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Excessive free radicals in human bodies are a leading cause of skin ageing and serious illnesses. To combat this issue, there is an increased interest in investigating dietary sources that are rich in antioxidants. The traditional Malaysian salad, known as *ulam*, has been used for decades by the locals for its potentials healing properties. However, there is a lack of scientific evidence regarding the antioxidant properties and carotenoid content of these *ulam* species, despite their high availability and economic feasibility. Therefore, this study aimed to investigate the potential of 40 different *ulam* species as sources of carotenoids and antioxidants. Our findings revealed that *ulam* is a rich, *halal* source of carotenoids and possesses significant antioxidant properties. Carotenoid levels differ among *ulam* species, with some exhibiting higher levels than others. The results showed that *ulam* can be recognised as a *halal* natural source of carotenoids for the food, cosmetic, and pharmaceutical industries. *Ulam* availability and economic feasibility make it a sustainable option for *halal* antioxidant supplementation. The findings of this study will benefit manufacturers, scientists, consumers, and others, with the potential to unlock the antioxidant potential of *ulam* and draw further attention to its nutritional and therapeutic benefits.

**Keywords:** *Ulam*; carotenoids; antioxidants; nutrition; *halal*

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Free radicals are molecules produced through cell metabolism and can naturally form in the body, playing a significant role in many normal cellular processes. Exposure to ionising radiation and toxic environments results in the production of free radicals in the body because of the death of cells caused by ionising radiation [1-3]. Environmental pollution, like heavy metal contamination and a high-oxygen atmosphere, can cause an increase in free radicals in the environment and stimulate cells in the body to generate free radicals [4, 5]. This occurs because free radicals contain oxygen elements that are commonly produced in living tissues and are known as reactive oxygen species (ROS) [6]. While ROS can be beneficial in moderate or low amounts, they can be harmful in high concentrations, leading to oxidative stress. Excessive concentrations of free radicals in bodies may destroy other molecules and cells, including lipids, proteins, DNA, and cell membranes, which are implicated in several human appearance changes such as pigmentation and the skin ageing

process, as well as serious diseases such as cancer, cardiovascular disease, and diabetes [7-10].

Antioxidants, or free radical scavengers, are chemical compounds that act as protective and neutralizing agents to prevent free radicals from causing harm. The body produces endogenous antioxidants, but in a small amount that functions to neutralise free radicals [11, 12]. Since this amount is not sufficient, the body needs to depend on external antioxidant sources. External antioxidant sources can be derived either from natural sources such as fruits, vegetables, herbs, spices, and grains or from artificial or synthetic sources. Some of the natural sources are also available as supplements, such as beta-carotene and vitamins A, C, and E [13]. Consuming fruits and vegetables is the simplest method of obtaining antioxidants, as they contain essential nutrients for growth and maintenance, along with bioactive compounds that can help slow down the aging process and prevent diseases [14-16].

Unfortunately, global industries predominantly use synthetic antioxidants because they are easy to obtain, inexpensive, stable during processing, and effective in suppressing oxidation [17]. The advantages of synthetic antioxidants are emphasised because of the lack of research on natural antioxidants, despite their potential to be safer than synthetic ones [18, 19]. Synthetic antioxidants such as butylated hydroxytoluene (BHT), butylated hydroxyanisole (BHA), pyrogallol (PY), and tert-butyl hydroquinone (TBHQ) are well-known and extensively used in various industries, such as food additives, cosmetics, and pharmaceuticals, including several healing products [20]. In the global era, consumers are increasingly aware of what they consume and use. Due to the potential cancer risks associated with long-term use of synthetic antioxidants, consumers are shifting their interest from synthetic to natural antioxidants [16, 21].

Malaysia is rich in traditional salads, with more than 120 species recognised within Asia. The old folks have consumed this vegetable, either raw or cooked, for decades. This traditional salad is also known as ulam. This diet has been practiced because they believe these ulam have several health benefits and can cure diseases [22]. In addition, ulam also consists of several medicinal functions such as antioxidant, antimicrobial, anti-inflammatory, neuroprotective, and others. It is also high in polyphenols, carotenoids, vitamins, and minerals [22, 23]. Studies by Islam et al. [23] revealed that differences in the composition of ulam contribute to the differences in antioxidant activities. Furthermore, some ulam is also recognised to contain several carotenoid compositions that function as the key to metabolism for human nutrition and health.

This study explores traditional Malaysian salad ulam as a potential natural source of carotenoids with antioxidant properties. Carotenoids are one of the most essential micronutrients and are an essential precursor of vitamin A that has been proven to ensure appropriate eye functions, and proper metabolic processes, and reduce the risk of several chronic illnesses [24, 25]. It has an essential antioxidant function in plants, which acts to deactivate the singlet oxygen that forms during photosynthesis [6, 26]. Fundamentally, carotenoids are produced by living organisms that undergo photosynthesis, especially plants, and come from non-photosynthetic organisms such as fungi. Carotenoids produced by fruits and leafy vegetables can be identified by their colour ranging from yellow-orange-red. For example, tomatoes are rich in lycopene, and turmeric is rich in curcumin [26].

The research focuses on the antioxidant potential of ulam, known for its diverse mix of herbs and vegetables. By investigating ulam as a *halal* carotenoid source, the study aims to highlight the health benefits of this traditional food item. This research contributes to the understanding of how traditional foods can serve as valuable sources of bioactive compounds that promote health and well-being. The

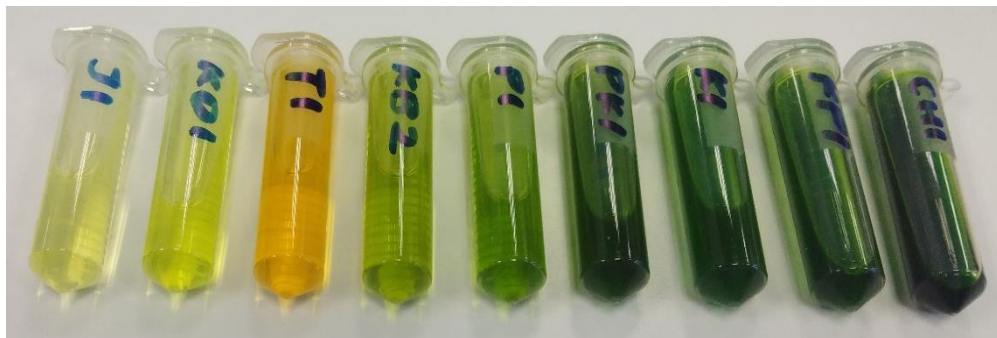
novelty also lies in the cultural significance of ulam within Malaysian cuisine. By highlighting ulam as a *halal* carotenoid source, the study bridges the gap between traditional culinary practices and modern nutritional science. This integration not only promotes the preservation of cultural heritage but also emphasises the importance of incorporating traditional foods into contemporary dietary patterns for enhanced health outcomes.

Ulam can be recognised as a *halal* natural source of carotenoids due to the benefits it offers, which comply with the regulations and standards set by authorities. According to the Malaysian Standard MS 1500:2019 [27], *Halal* Food – General Requirement which governs the production, preparation, handling, and storage of *halal* food, under item 4.5.1.2, it is generally accepted that all plant-based products and derivatives are *halal*, except for those that are harmful or pose a health risk. Similarly, in the pharmaceutical sector, the Malaysian Standard MS 2424:2012 [28] delineates comprehensive guidelines to produce *halal* pharmaceutical products. One important requirement specified in this standard is that the plants used in manufacturing must be non-prohibited species according to regulatory and national laws. In the realm of *halal* food, it is crucial to verify that all ingredients, including those derived from plants, adhere to Islamic dietary laws [29, 30]. The restriction of specific plant species in *halal* products is determined by Islamic dietary laws, which distinguish between what is allowed (*halal*) and what is prohibited (*haram*) for consumption [31]. Regulatory bodies overseeing *halal* food production typically maintain a list of plant species that are forbidden in *halal* products. This list can be used by producers as a guide to ensure compliance with *halal* standards. By aligning with both *halal* standards and national regulations, producers can guarantee that their products meet the required criteria for quality, safety, and genuineness [32-34]. Therefore, this study will propose alternatives for *halal* natural carotenoids, focusing on the antioxidant potential of ulam.

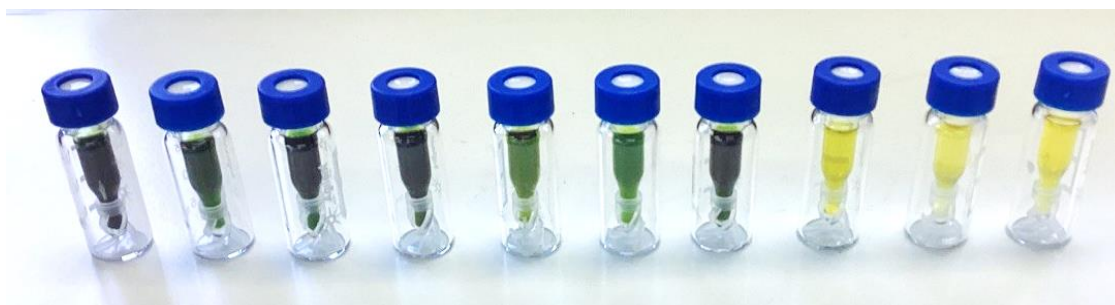
## EXPERIMENTAL

### Analysis of Carotenoid Composition

A total of 40 samples of selected ulam were collected from the wet market in Selayang, Selangor. Samples were then washed, chopped, and labelled before being inserted into the freeze-dryer and run for 72 hours to remove water content. Afterward, samples were ground into powder, kept in an airtight container, and placed in a dark room. Next, the extraction of carotenoid pigments was done by using the alkaline method. Each sample was weighed with 1.0 g of dried weight, added to a sodium hydroxide solution, followed by an ethyl acetate solution, and left overnight for 24 hours at a temperature of -20 °C. Subsequently, the extracted pigment was collected, kept in an Eppendorf tube, and stored at -20 °C prior to further analysis (Figure 1) [35].



**Figure 1.** The extracted carotenoid pigments from ulam were kept in an Eppendorf tube and stored at  $-20^{\circ}\text{C}$  prior to further processing.



**Figure 2.** The extracted carotenoid samples were prepared in HPLC vials.

The analysis of carotenoid content and composition was conducted using high-performance liquid chromatography (HPLC). 300  $\mu\text{L}$  of the extracted carotenoid sample was inserted into a vial and prepared in three replicates (Figure 2). Each replicate of the sample took 40 minutes to be analysed by HPLC [36].

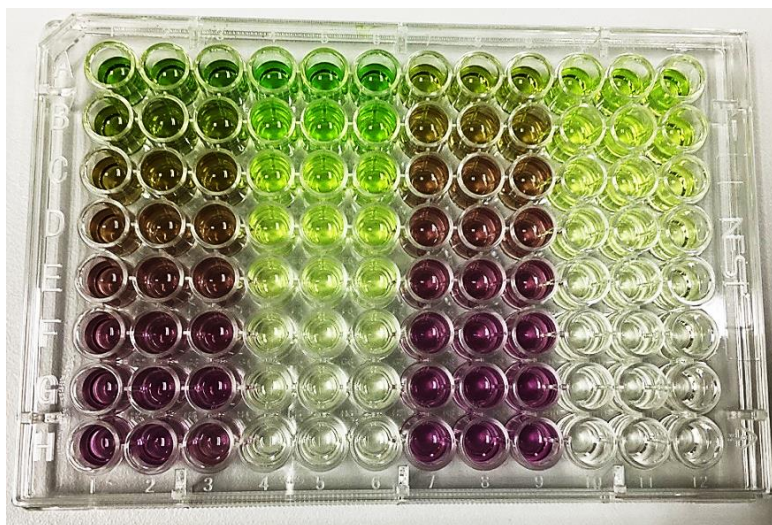
#### DPPH Free Radical Scavenging Assay

2,2-Diphenyl-1-picrylhydrazyl (DPPH) from Sigma-Aldrich was used to test free radical scavenging in the top three ulam samples. Three extracts from the ulam with the highest carotenoid content were chosen for this assay. The extract was dissolved in methanol (MeOH) to produce a stock solution with a concentration of 1  $\mu\text{g}/\mu\text{L}$ . Approximately 100  $\mu\text{L}$  of MeOH was added using a multipipettor (8 tips) into a 96-well microplate from well B1 to B6, continually

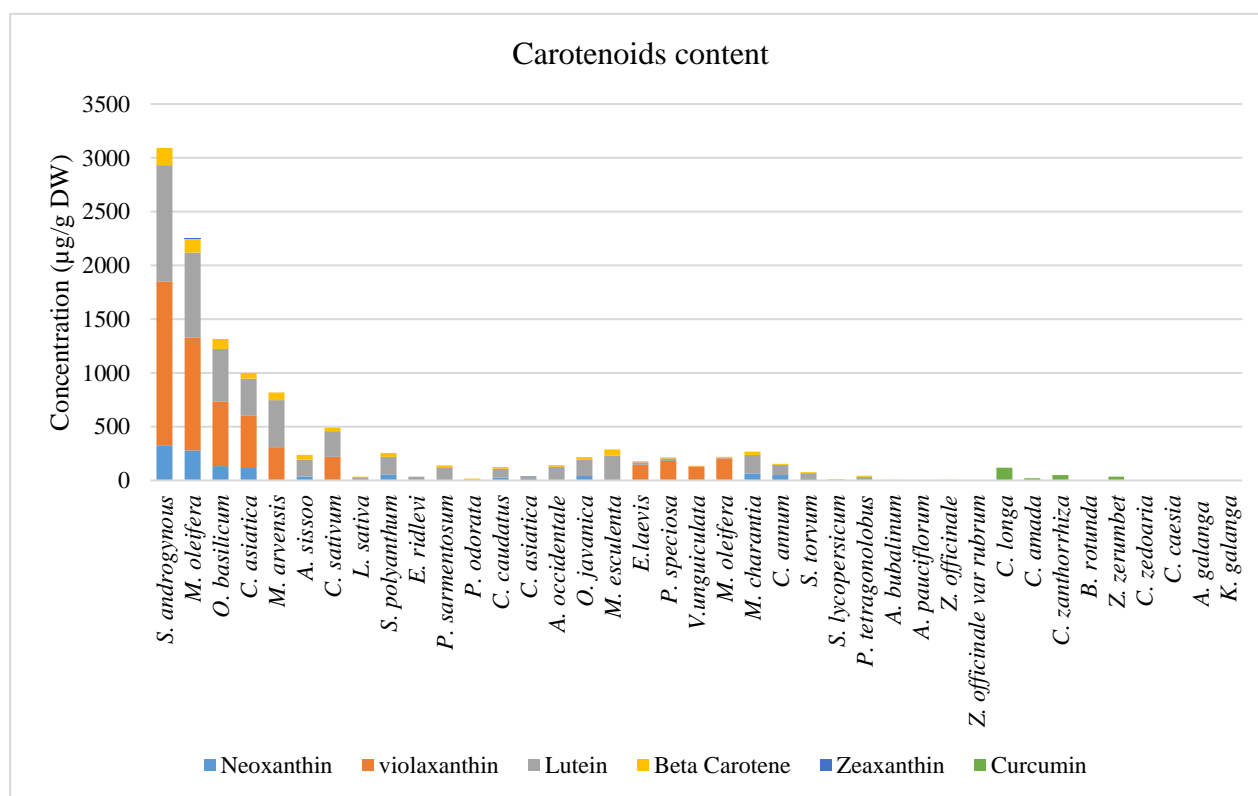
until the last row (H1-H6). Next, 200  $\mu\text{L}$  of the prepared sample extract was pipetted into wells A1 to A6. All samples in the first row (A1 to A6) were then transferred to the well in row B and mixed properly using a pipette 4–5 times before being two-fold diluted until row H. Then, 100  $\mu\text{L}$  of MeOH was added to A4 to A6 until the last row of H4 to H6, respectively. 100  $\mu\text{L}$  of DPPH methanolic solution was added to A1 to A3 until the last row of H1 to H3, respectively (Figure 3). The final concentrations of the sample were 500, 250, 125, 62.5, 31.3, 15.6, 7.8, and 3.9  $\mu\text{g}/\text{mL}$ . Finally, the microplate was covered with a lid and kept in the dark at room temperature for 40 minutes. After that, absorbance was read at 517 nm of wavelength using a Varioskan™ LUX multimode microplate reader (Thermo Scientific™). The result obtained was calculated and expressed as the percentage of DPPH free radical scavenging activity using the formula stated below [37]:

$$\frac{1 - \text{Absorbance sample}}{\text{Absorbance DPPH}} \times 100 = \% \text{ Inhibition}$$

\*Absorbance sample = Absorbance (sample + DPPH) – Absorbance (sample+ MeOH)



**Figure 3.** The extracted samples were prepared for antioxidant DPPH analysis.



**Figure 4.** Carotenoids content of 40 ulam species

## RESULTS AND DISCUSSION

### Analysis of Carotenoid Composition

The HPLC analysis results showed that the total carotenoid content in all samples ranged between  $1.14 \pm 0.02$  and  $1078.30 \pm 3.60$   $\mu\text{g/g DW}$  (Figure 4).

The three samples with the highest lutein,  $\beta$ -carotene, and violaxanthin were *Sauropus androgynus* (Cekur manis) ( $1078.30 \pm 3.60$ ,  $160.17 \pm 3.85$ , and  $1525.30 \pm 13.38$   $\mu\text{g/g DW}$ ), *Moringa oleifera* (Kelor leaves) ( $793.26 \pm 8.97$ ,  $122.92 \pm 8.30$ , and  $1048.60 \pm 11.58$   $\mu\text{g/g DW}$ ), and *Ocimum basilicum* (Selasih) ( $493.03 \pm 8.32$ ,  $90.68 \pm 0.93$ , and  $598.55 \pm 24.95$   $\mu\text{g/g DW}$ ) (Figure 5). Other

carotenoids such as neoxanthin, zeaxanthin, and curcumin were also detected in several samples analysed, but with lower content. The highest amount of carotenoids in these three samples is supported by previous reports. *S. androgynus* was identified as having the highest total carotenoid content in the study by Othman et al. [38] and Andarwulan et al. [39]. In addition, Platel and Srinivasan [40] reported that carotenoid content in *S. androgynus* depends on the maturity of the leaves. As for *M. oleifera*, higher  $\beta$ -carotene was found in the dried leaves than in its foliage, fruits, and flowers [41, 42]. While other carotenoids such as lutein, violaxanthin, and zeaxanthin were also detected, but in lesser amounts [43]. The study by Dumbra et al. [44] showed that *O. basilicum* extracted with two different methods had the highest carotenoid composition compared to other vegetables tested. This result was supported by Filip [45], who reported that *O. basilicum* contains high concentrations of carotenoids, with  $\beta$ -carotene being the main compound detected. A previous study also detected high concentrations of violaxanthin and  $\beta$ -carotene in *O. basilicum*, with levels of  $977 \pm 15.56$  and  $204.75 \pm 1.77$   $\mu\text{g/g}$  DW, respectively [46]. These studies also noted that  $\beta$ -carotene offers more benefits than vitamin A alone and is recognised as a potent antioxidant. Consequently, the three samples with the highest carotenoid content were selected for further antioxidant assays.

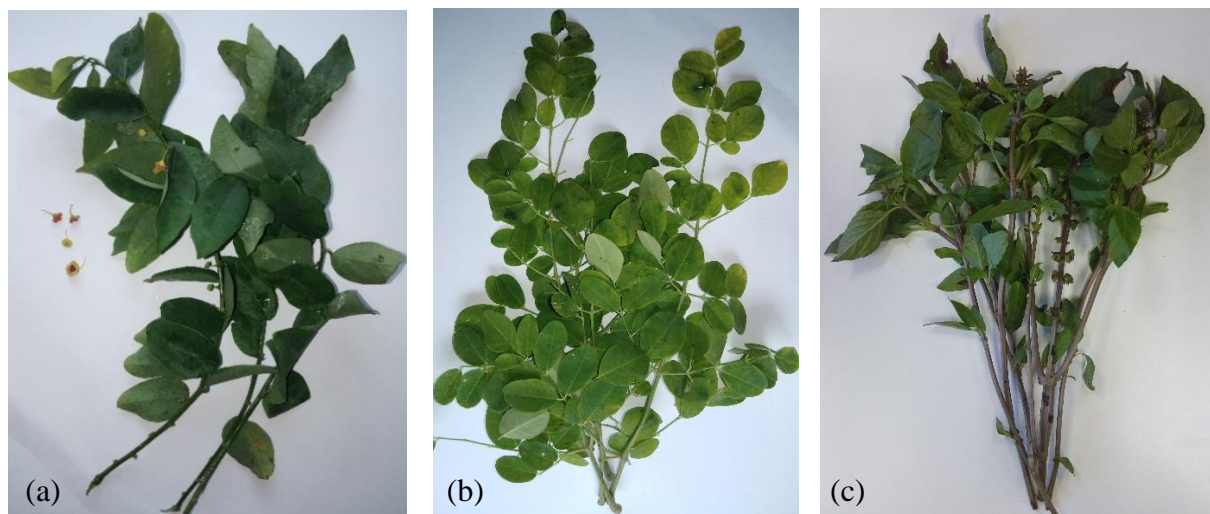
### Antioxidant Activity

The Varioskan™ LUX multimode microplate reader (Thermo Scientific™) at wavelength 517 nm was used to analyse the antioxidant activity of the three ulam samples. The inhibition percentage for ascorbic acid standard, *S. androgynus* (Cekur manis), *M. oleifera* (Kelor leaves), and *O. basilicum* (Selasih) is

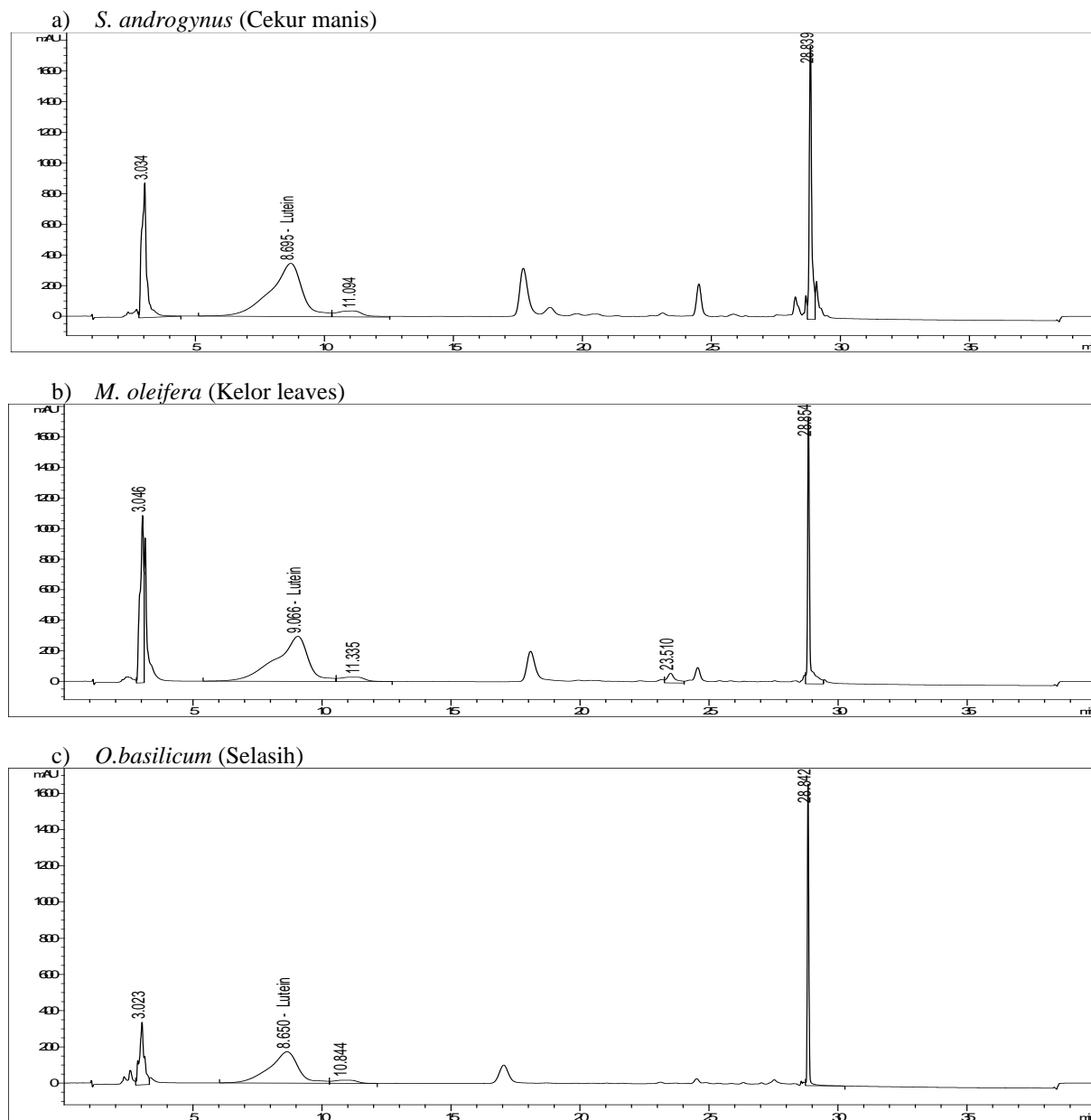
demonstrated in Figure 7. From the graph, it showed that *S. androgynus* has exhibited a higher DPPH scavenging activity with 98% inhibition at a concentration of 500  $\mu\text{g/mL}$  compared to *M. oleifera* and *O. basilicum*, with 91% and 89% inhibition at a concentration of 500  $\mu\text{g/mL}$ , respectively.

The scavenging activity of *S. androgynus* is comparable to the findings of Kuttinath et al. [47], which detected more than 70% DPPH inhibition at a concentration of 100  $\mu\text{g/mL}$ . Other studies have also noted that this ulam contains significant amounts of phenolic compounds, carotenoids, antioxidants, vitamins, and minerals that contribute to its therapeutic properties [48]. Kumar et al. [49] reported excellent scavenging activity of *M. oleifera* with 95.52% inhibition at a concentration of 100  $\mu\text{g/mL}$ , while Siti Mahirah et al. [50] demonstrated a DPPH scavenging activity of 92.60% for *O. basilicum* at the same concentration. Naidu et al. [51] found 73.75% DPPH inhibition for the same ulam species, highlighting *O. basilicum*'s high  $\beta$ -carotene content, which contributes to its potent antioxidant capacity. Several studies have also reported positive scavenging activities of *S. androgynus*, *M. oleifera*, and *O. basilicum*, highlighting their potential as powerful antioxidant agents [38, 52, 53, 54]. However, variations in inhibition percentages may arise due to factors such as plant maturity, temperature, pH, storage duration, solvent type, and extraction method [54, 55].

The findings of this research support the exceptional qualities of *S. androgynus*, *M. oleifera*, and *O. basilicum* as plants with diverse applications in traditional ethnic cuisines and ethnomedicinal traditions [56-58]. Given their high antioxidant levels, these ulam species have the potential to protect cells from oxidative damage and serve as precursors to vitamin A.



**Figure 5.** Three ulam species with the highest carotenoids content; a) *S. androgynus* (Cekur manis), b) *M. oleifera* (Kelor leaves), c) *O. basilicum* (Selasih).

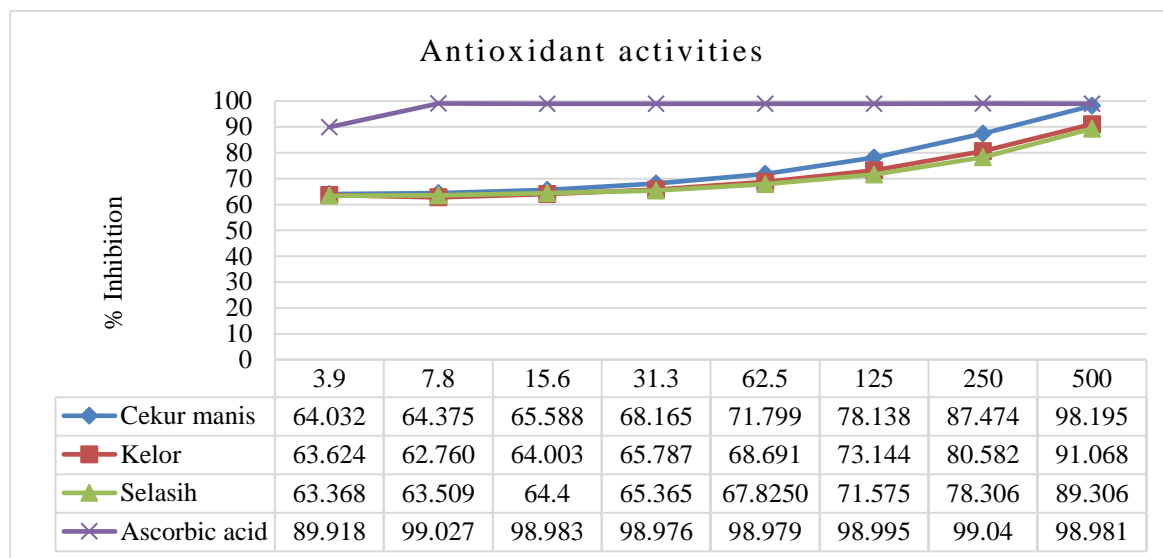


**Figure 6.** HPLC chromatogram of three ulam species showing the highest content and composition of carotenoids: lutein,  $\beta$ -carotene, and violaxanthin.

Carotenoids, renowned for their potent antioxidant properties, help promote overall well-being by combating harmful free radicals and reducing oxidative stress [59]. Beyond dietary significance, ulam shows promise for future medicinal applications based on emerging research [60-63].

Furthermore, numerous studies have identified a variety of phytochemicals in ulam beyond antioxidants, suggesting their potential medicinal properties [64-66]. Indigenous communities have long utilized ulam for its medicinal properties in treating common ailments [67, 68], and current research, including this study, aims to scientifically validate these traditional uses. Continued exploration of ulam's phytochemical

composition may reveal new bioactive compounds with therapeutic potential against various diseases, potentially leading to the development of novel medicines [57, 69, 70]. Therefore, as highlighted in this study, the presence of antioxidants in ulam not only enhances dietary diversity but also presents a range of bioactive compounds with potential medicinal uses, deserving further exploration and research [71]. Studying the carotenoid content and composition of ulam is crucial for advancing the pharmaceutical industry and the global market by discovering alternative *halal* active pharmaceutical ingredients [46]. Findings on ulam with the highest carotenoid content can be further explored using green technology, benefiting Malaysia's *halal* pharmaceutical sector.



**Figure 7.** Antioxidant activities (inhibition percentage) of *S. androgynus* (Cekur manis), *M. oleifera* (Kelor leaves), and *O. basilicum* (Selasih) as well as the ascorbic acid standard.

## CONCLUSION

Given the potential links between synthetic antioxidants and cancer, there is a growing shift in the food, cosmetics, and pharmaceutical industries towards natural and safe antioxidant sources derived from plants, renowned for their potent antioxidant properties. Moreover, with the exception of those prohibited by Islamic authorities, all plant species, their products, and byproducts are considered *halal*. This research aims to identify optimal sources of natural antioxidants. *Sauropus androgynus*, *Moringa oleifera*, and *Ocimum basilicum* are highlighted as promising natural antioxidant sources due to their high carotenoid content. The robust biological activity of carotenoids supports diverse commercial applications and contributes to the growth of the *halal* market. Further investigations are required to substantiate the potential of ulam as a *halal* active pharmaceutical ingredient. Consequently, this discovery is expected to benefit society and generate interest among producers, researchers, consumers, and other stakeholders committed to advancing human health.

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## REFERENCES

- Jiao, Y., Cao, F. and Liu, H. (2022) Radiation-induced cell death and its mechanisms. *Health Physics*, **123**(5), 376–386.
- Lumniczky, K., Impens, N., Armengol, G., Candéias, S., Georgakilas, A. G., Hornhardt, S., Martin, O. A., Rödel, F. and Schaeue, D. (2021) Low dose ionizing radiation effects the immune system. *Environment International*, **149**, 106212.
- Valko, M., Leibfritz, D., Moncol, J., Cronin, M. T., Mazur, M. and Telser, J. (2007) Free radicals and antioxidants in normal physiological functions and human disease. *The International Journal of Biochemistry and Cell Biology*, **39**(1), 44–84.
- Atasoy, N. and Yücel, U. M. (2021) Antioxidants from plant sources and free radicals. In *Reactive Oxygen Species*, eds. R. Ahmad, IntechOpen, London.
- Tripathi, R., Gupta, R., Sahu, M., Srivastava, D., Das, A., Ambasta, R. K. and Kumar, P. (2022) Free radical biology in neurological manifestations: mechanisms to therapeutics interventions. *Environmental Science and Pollution Research*, **29**, 62160–62207.
- Sharma, R. and Ghoshal, G. (2020) Optimization of carotenoids production by *Rhodotorula mucilaginosa* (MTCC-1403) using agro-industrial waste in bioreactor: A statistical approach. *Biotechnology Reports*, **25**, e00407.
- Adetuyi, B. O., Adebayo, P. F., Olajide, P. A., Atanda, O. O. and Oloke, J. K. (2022) Involvement of free radicals in the ageing of cutaneous membrane. *World News of Natural Sciences*, **43**, 11–37.

8. Ajayi, E. I. O., Adeleke, M. A., Adewumi, T. Y. and Adeyemi, A. A. (2017) Antiplasmodial activities of ethanol extracts of *Euphorbia hirta* whole plant and *Vernonia amygdalina* leaves in *Plasmodium berghei*-infected mice. *Journal of Taibah University for Science*, **11**(6), 831–835.
9. Juan, C. A., Pérez de la Lastra, J. M., Plou, F. J. and Pérez-Lebeña, E. (2021) The chemistry of reactive oxygen species (ROS) revisited: outlining their role in biological macromolecules (DNA, lipids and proteins) and induced pathologies. *International Journal of Molecular Sciences*, **22**(9), 4642.
10. Sadiq, I. Z. (2023) Free radicals and oxidative stress: Signaling mechanisms, redox basis for human diseases, and cell cycle regulation. *Current Molecular Medicine*, **23**(1), 13–35.
11. Martemucci, G., Costagliola, C., Mariano, M., D'andrea, L., Napolitano, P. and D'Alessandro, A. G. (2022) Free radical properties, source and targets, antioxidant consumption and health. *Oxygen*, **2**(2), 48–78.
12. Mirończuk-Chodakowska, I., Witkowska, A. M. and Zujko, M. E. (2018) Endogenous non-enzymatic antioxidants in the human body. *Advances in Medical Sciences*, **63**(1), 68–78.
13. Bouayed, J. and Bohn, T. (2010) Exogenous antioxidants—double-edged swords in cellular redox state: health beneficial effects at physiologic doses versus deleterious effects at high doses. *Oxidative Medicine and Cellular Longevity*, **3**(4), 228–237.
14. Arias, A., Feijoo, G. and Moreira, M. T. (2022) Exploring the potential of antioxidants from fruits and vegetables and strategies for their recovery. *Innovative Food Science and Emerging Technologies*, **77**, 102974.
15. Jideani, A. I., Silungwe, H., Takalani, T., Omolola, A. O., Udeh, H. O. and Anyasi, T. A. (2021) Antioxidant-rich natural fruit and vegetable products and human health. *International Journal of Food Properties*, **24**(1), 41–67.
16. Tanvir, E. M., Hossen, M., Hossain, M., Afroz, R., Gan, S. H., Khalil, M. and Karim, N. (2017) Antioxidant properties of popular turmeric (*Curcuma longa*) varieties from Bangladesh. *Journal of Food Quality*, **2017**(1), 8471785.
17. Makahleh, A., Saad, B. and Bari, M. F. (2015) Synthetic phenolics as antioxidants for food preservation. In *Handbook of Antioxidants for Food Preservation*, eds. F. Shahidi, Woodhead Publishing, San Francisco.
18. Stoia, M. and Oancea, S. (2022) Low-molecular-weight synthetic antioxidants: classification, pharmacological profile, effectiveness and trends. *Antioxidants*, **11**(4), 638.
19. Pokorný, J. (2007) Are natural antioxidants better—and safer—than synthetic antioxidants? *European Journal of Lipid Science and Technology*, **109**(6), 629–642.
20. Oladele, J. O., Ajayi, E. I., Oyeleke, O. M., Oladele, O. T., Olowookere, B. D., Adeniyi, B. M. and Oladiji, A. T. (2020) A systematic review on COVID-19 pandemic with special emphasis on curative potentials of Nigeria based medicinal plants. *Heliyon*, **6**(9), e04897.
21. Mitterer-Daltoé, M., Bordim, J., Lise, C., Breda, L., Casagrande, M. and Lima, V. (2020) Consumer awareness of food antioxidants. Synthetic vs. Natural. *Food Science and Technology*, **41**, 208-212.
22. You, Y., Shahar, S., Haron, H. and Yahya, H. (2018) More ulam for your brain: A review on the potential role of ulam in protecting against cognitive decline. *Sains Malaysiana*, **47**(11), 2713–2729.
23. Islam, R., Uddin, M. N., Ashrafuzzaman, M. and Hoque, M. I. U. (2018) Phenolics and carotenoids contents and radical scavenging capacity of some selected solanaceous medicinal plants. *Journal of the Bangladesh Agricultural University*, **16**(1), 56–61.
24. Gebregziabher, B. S., Gebremeskel, H., Debesa, B., Ayalneh, D., Mitiku, T., Wendwessen, T., Habtemariam, E., Nur, S. and Getachew, T. (2023) Carotenoids: Dietary sources, health functions, biofortification, marketing trend and affecting factors—A review. *Journal of Agriculture and Food Research*, **14**, 100834.
25. Gürbüz, M. and Aktaş, Ş. (2022) Understanding the role of vitamin A and its precursors in the immune system. *Nutrition Clinique et Métabolisme*, **36**(2), 89–98.
26. Bohn, T. (2017) Bioactivity of carotenoids—chasm of knowledge. *International Journal for Vitamin and Nutrition Research*, **10**, 1–5.
27. Malaysian Standard MS 1500 (2019) Halal Food – General Requirement (Third version). *Department of Standard Malaysia: Cyberjaya*.
28. Malaysia Standard MS 2424 (2012) Halal Pharmaceuticals - General guidelines. *Department of Standard Malaysia: Cyberjaya*.



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29. Asa, R. S. and Azmi, I. M. A. G. (2018) The concept of halal and halal food certification process in Malaysia: Issues and concerns. *Malaysian Journal of Consumer and Family Economics*, **20**, 38–50.
30. Tahir, P. and Muslih, M. (2023) Halal and Safe Food in Islamic Law. *Batulis Civil Law Review*, **4(1)**, 37–50.
31. Mokti, H. A. and Balwi, M. A. W. F. M. (2022) Halal Food Quality: An Analysis of Relevant Guidelines and Regulations in Malaysia. *Journal of Fatwa Management and Research*, **27(2-SE)**, 37–55.
32. Ahmad, Z., Mafaz, M. N. A. and Rahman, M. M. (2023) Harmony in halal: Understanding stakeholder views analyzing products and evaluating policies in Malaysia. *West Science Business and Management*, **1(05)**, 495–508.
33. Al-Shami, H. A. and Abdullah, S. (2023) Halal food industry certification and operation challenges and manufacturing execution system opportunities. A review study from Malaysia. *Materials Today: Proceedings*, **80**, 3607–3614.
34. Mohd Hatta, F. A., Mat Ali, Q. A., Mohd Kashim, M. I. A., Othman, R., Abd Mutalib, S. and Mohd Nor, N. H. (2023) Recent advances in halal bioactive materials for intelligent food packaging indicator. *Foods*, **12(12)**, 2387.
35. Hatta, F. A. M. and Othman, R. (2020) Carotenoids as potential biocolorants: A case study of astaxanthin recovered from shrimp waste. In *Carotenoids: Properties, processing and applications*, eds. C. M. Galanakis, Academic Press, London.
36. Othman, R. (2009) Biochemistry and genetics of carotenoid composition in potato tubers. *Ph.D. Thesis, Lincoln University, Christchurch, New Zealand*.
37. Hatta, F. A. M. (2019) Study on lipophilic (C40) and hydrophilic (C50) carotenoids: production from shrimp waste and saffron as potential halal pigments. *Ph.D. Thesis, International Islamic University Malaysia, Kuala Lumpur, Malaysia*.
38. Othman, R., Zaifuddin, F. A. M. and Hassan, N. M. (2017) Characterisation of carotenoid and total retinol equivalent content in Ulam and medicinal species as alternative food intervention to combat vitamin A deficiency. *Journal of Pharmacy and Nutrition Sciences*, **7(3)**, 81–87.
39. Andarwulan, N., Kurniasih, D., Apriady, R. A., Rahmat, H., Roto, A. V. and Bolling, B. W. (2012) Polyphenols, carotenoids, and ascorbic acid in underutilized medicinal vegetables. *Journal of Functional Foods*, **4(1)**, 339–347.
40. Platel, K. and Srinivasan, K. (2017) Nutritional profile of chekurmanis (*Sauropus androgynus*), A less explored green leafy vegetable. *Indian Journal of Nutrition and Dietetics*, **54**, 243–352.
41. Glover-Amengor, M., Aryeetey, R., Afari, E. and Nyarko, A. (2017) Micronutrient composition and acceptability of *Moringa oleifera* leaf-fortified dishes by children in Ada-East district, Ghana. *Food Science and Nutrition*, **5(2)**, 317–323.
42. Saini, R. K., Shetty, N. P. and Giridhar, P. (2014) Carotenoid content in vegetative and reproductive parts of commercially grown *Moringa oleifera* Lam. cultivars from India by LC–APCI–MS. *European Food Research and Technology*, **238(6)**, 971–978.
43. Ma, Z. F., Ahmad, J., Zhang, H., Khan, I. and Muhammad, S. (2020) Evaluation of phytochemical and medicinal properties of *Moringa oleifera* as a potential functional food. *South African Journal of Botany*, **129**, 40–46.
44. Dumbrava, D. G., Moldovan, C., Raba, D. N. and Popa, M. V. (2012) Vitamin C, chlorophylls, carotenoids and xanthophylls content in some basil (*Ocimum basilicum* L.) and rosemary (*Rosmarinus officinalis* L.) leaves extracts. *Journal of Agrolimentary Processes and Technologies*, **18(3)**, 253–258.
45. Filip, S. (2017) Basil (*Ocimum basilicum* L.) a source of valuable phytonutrients. *International Journal of Clinical Nutrition & Dietetics*, **3**, 118.
46. Mat Ali, Q. A., Mohd Hatta, F. A., Othman, R., Ramya, R., Mohd Latiff, N. H. & Wan Sulaiman, W. S. H. (2023) Carotenoid composition in twenty ulam species as potential halal active pharmaceutical ingredients. *Food Research*, **7(4)**, 331–336.
47. Kuttinath, S., Kh, H. and Rammohan, R. (2019) Phytochemical screening, antioxidant, antimicrobial, and antibiofilm activity of *Sauropus androgynus* leaf extracts. *Asian Journal of Pharmaceutical and Clinical Research*, **12(4)**, 244–250.
48. Eng Khoo, H., Azlan, A. and Ismaila, A. (2015) *Sauropus androgynus* leaves for health benefits: hype and the science. *The Natural Products Journal*, **5(2)**, 115–123.
49. Kumar, V., Pandey, N., Mohan, N. and Singh, R. P. (2012) Antibacterial and antioxidant activity of different extract of *Moringa oleifera* Leaves—an in vitro study. *International Journal of*

- Pharmaceutical Sciences Review and Research*, **12(1)**, 89–94.
50. Siti Mahirah, Y., Rabeta, M. S. and Antora, R. A. (2018) Effects of different drying methods on the proximate composition and antioxidant activities of *Ocimum basilicum* leaves. *Food Research*, **2(5)**, 421–428.
51. Naidu, J. R., Ismail, R. B. and Sasidharan, S. (2016) Chemical profiling and antioxidant activity of Thai basil (*Ocimum basilicum*). *Journal of Essential Oil Bearing Plants*, **19(3)**, 750–755.
52. Norhayati, Y., Nurulhidayah, A., Rini Zunnurni, M. J., Norliana, A. R., Norliana, W. and Mohd Ifwat, I. (2018) Antioxidative constituents of selected Malaysian 'Ulam'. *Science International*, **30**, 273–278.
53. Bunawan, H., Bunawan, S. N., Baharum, S. N. and Noor, N. M. (2015) *Sauropus androgynus* (L.) Merr. induced bronchiolitis obliterans: from botanical studies to toxicology. *Evidence-Based Complementary and Alternative Medicine*, **2015**, 714158.
54. Hikmawanti, N. P. E., Fatmawati, S. and Asri, A. W. (2021) The effect of ethanol concentrations as the extraction solvent on antioxidant activity of Katuk (*Sauropus androgynus* (L.) Merr.) leaves extracts. *IOP Conference Series: Earth and Environmental Science*, **755**, 012060.
55. Arabshahi-Delouee, S. and Urooj, A. (2007) Antioxidant properties of various solvent extracts of mulberry (*Morus indica* L.) leaves. *Food chemistry*, **102(4)**, 1233–1240.
56. Arora, S. and Arora, S. (2021) Nutritional significance and therapeutic potential of *Moringa oleifera*: The wonder plant. *Journal of Food Biochemistry*, **45(10)**, e13933.
57. Jamal, J. A. and Husain, K. (2022) Traditional Malay ulam for healthy ageing. In *Healthy Ageing in Asia*, eds. G. C. Soon, G. Bodeker, K. Kariippanon, CRC Press, Boca Raton.
58. Romano, R., De Luca, L., Aiello, A., Pagano, R., Di Pierro, P., Pizzolongo, F. and Masi, P. (2022) Basil (*Ocimum basilicum* L.) leaves as a source of bioactive compounds. *Foods*, **11(20)**, 3212.
59. Mohd Hatta, F. A., Othman, R., Mat Ali, Q. A., Mohd Hassan, N., Ramya, R., Wan Sulaiman, W. S., Mohd Latiff, N. H. & Mohd Kashim, M. I. A. (2023) Carotenoids composition, antioxidant, and antimicrobial capacities of *Crocus sativus* L. stigma. *Food Research*, **7(4)**, 337–343.
60. Anju, T., Rai, N. K. S. and Kumar, A. (2022) *Sauropus androgynus* (L.) Merr.: a multipurpose plant with multiple uses in traditional ethnic culinary and ethnomedicinal preparations. *Journal of Ethnic Foods*, **9(1)**, 10.
61. Muteeb, G., Aatif, M., Farhan, M., Alsultan, A., Alshoaibi, A. and Alam, M. W. (2023) Leaves of *Moringa oleifera* are potential source of bioactive compound  $\beta$ -carotene: Evidence from in silico and quantitative gene expression analysis. *Molecules*, **28(4)**, 1578.
62. Nadeem, H. R., Akhtar, S., Sestili, P., Ismail, T., Neugart, S., Qamar, M. and Esatbeyoglu, T. (2022) Toxicity, antioxidant activity, and phytochemicals of basil (*Ocimum basilicum* L.) leaves cultivated in Southern Punjab, Pakistan. *Foods*, **11(9)**, 1239.
63. Noor, Z. I., Ahmed, D., Rehman, H. M., Qamar, M. T., Froeyen, M., Ahmad, S. and Mirza, M. U. (2019) In vitro antidiabetic, anti-obesity and antioxidant analysis of *Ocimum basilicum* aerial biomass and in silico molecular docking simulations with  $\alpha$ -amylase and lipase enzymes. *Biology*, **8(4)**, 92.
64. Chhikara, N., Kaur, A., Mann, S., Garg, M. K., Sofi, S. A. and Panghal, A. (2021) Bioactive compounds associated health benefits and safety considerations of *Moringa oleifera* L.: An updated review. *Nutrition and Food Science*, **51(2)**, 255–277.
65. Mazumdar, P., Jalaluddin, N. S. M., Nair, I., Tian Tian, T., Rejab, N. A. B. and Harikrishna, J. A. (2023) A review of *Hydrocotyle bonariensis*, a promising functional food and source of health-related phytochemicals. *Journal of Food Science and Technology*, **60(10)**, 2503–2516.
66. Roy, S. and Chaudhuri, T. K. (2020) A comprehensive review on the pharmacological properties of *Diplazium esculentum*, an edible fern. *Journal of Pharmacy and Pharmacology*, **3**, 1–9.
67. Madjos, G. G. and Ramos, K. P. (2021) Ethnobotanical indigenous knowledge, systems and practices (IKSP) in Western Mindanao, Philippines: The sea nomads “Bajau” of Zamboanga City. *International Journal of Botany Studies*, **6(5)**, 819–825.
68. Tharmabalan, R. T. (2023) Nutritional profiles of four promising wild edible plants commonly consumed by the Semai in Malaysia. *Current Developments in Nutrition*, **7(4)**, 100054.
69. Arif, T. (2020) Therapeutic potential and traditional uses of *Sauropus androgynus*: a review. *Journal of Pharmacognosy and Phytochemistry*, **9(3)**, 2131–2137.

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70. Zhang, B. D., Cheng, J. X., Zhang, C. F., Bai, Y. D., Liu, W. Y., Li, W., Koike, K., Akihisa, T., Feng, F. and Zhang, J. (2020) *Sauropus androgynus* L. Merr. -A phytochemical, pharmacological and toxicological review. *Journal of Ethnopharmacology*, **257**, 112778.

71. Amir Rawa, M. S., Mazlan, M. K. N., Ahmad, R., Nogawa, T. and Wahab, H. A. (2022) Roles of *Syzygium* in anti-cholinesterase, anti-diabetic, anti-inflammatory, and antioxidant: from Alzheimer's perspective. *Plants*, **11(11)**, 1476.